

## CHAPTER 10

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# Histamine in Neurotransmission and Brain Diseases

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### Abstract

**A**part from its central role in the mediation of allergic reactions, gastric acid secretion and inflammation in the periphery, histamine serves an important function as a neurotransmitter in the central nervous system. The histaminergic neurons originate from the tuberomammillary nucleus of the posterior hypothalamus and send projections to most parts of the brain. The central histamine system is involved in many brain functions such as arousal, control of pituitary hormone secretion, suppression of eating and cognitive functions. The effects of neuronal histamine are mediated via G-protein-coupled H<sub>1</sub>-H<sub>4</sub> receptors. The prominent role of histamine as a wake-promoting substance has drawn interest to treat sleep-wake disorders, especially narcolepsy, via modulation of H<sub>3</sub> receptor function. Post mortem studies have revealed alterations in histaminergic system in neurological and psychiatric diseases. Brain histamine levels are decreased in Alzheimer's disease patients whereas abnormally high histamine concentrations are found in the brains of Parkinson's disease and schizophrenic patients. Low histamine levels are associated with convulsions and seizures. The release of histamine is altered in response to different types of brain injury: e.g. increased release of histamine in an ischemic brain trauma might have a role in the recovery from neuronal damage. Neuronal histamine is also involved in the pain perception. Drugs that increase brain and spinal histamine concentrations have antinociceptive properties. Histaminergic drugs, most importantly histamine H<sub>3</sub> receptors ligands, have shown efficacy in many animal models of the above-mentioned disorders. Ongoing clinical trials will reveal the efficacy and safety of these drugs in the treatment of human patients.

### Histaminergic Neurons

The first findings of histamine in the brain date back to 1919 when John J. Abel isolated histamine from the pituitary.<sup>1</sup> However, histamine's role as a neurotransmitter became evident only several decades later when lesions of the lateral hypothalamic area were found to decrease the activity of histamine synthesizing enzyme, L-histidine decarboxylase (HDC).<sup>2</sup> Another decade went by before methods became available to directly demonstrate the localization of the histaminergic neurons in the brain.<sup>3,4</sup> Cell bodies of histaminergic neurons are localized in the tuberomammillary nucleus (TMN) of the posterior hypothalamus from where they send projections to essentially all areas of the central nervous system similar to other amines (Fig. 1).<sup>5</sup> The number of histamine-containing neurons is about 4000 in the rat<sup>6</sup> whereas in human brain histaminergic neurons are more numerous (>64,000).<sup>7</sup>

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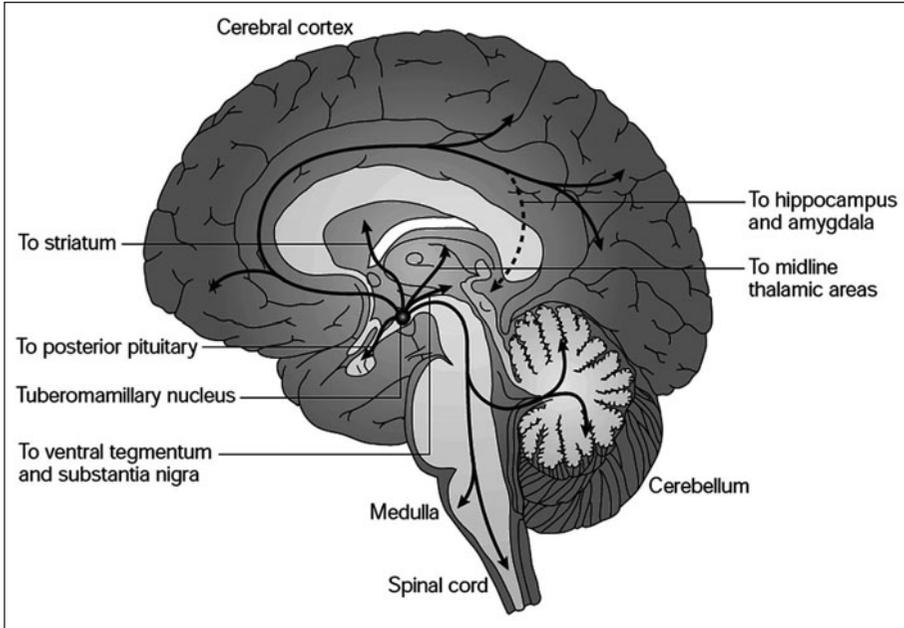


Figure 1. Histaminergic neurons and main projections in the human brain. Reprinted from: Haas H, Panula P. *Nat Rev Neurosci* 2003; 4:121-130.<sup>5</sup>

In addition to neurons, mast cells can produce histamine in the brain.<sup>8</sup> Indications of histamine synthesis in microglial cells also exists, but *in vivo* evidence of histamine produced by microglia is still missing.<sup>9</sup> It is noteworthy that brain ependymal cells also express HDC and can potentially synthesize histamine.<sup>10</sup> The role of this histamine is unknown, but it may be involved in regulation of stem cells located underneath the ependymal layer. Neural stem cells *in vitro* respond to both H<sub>1</sub>R and H<sub>2</sub>R receptor ligands.<sup>11</sup> Nonneuronal histamine has an important role in immune responses in the periphery and in the CNS and is covered elsewhere in this book. However, it should be noted that the source of brain histamine in some cases is difficult to detect. It is thus possible that both neuronal and nonneuronal histamine might regulate certain brain functions such as neuroinflammation.

### Histamine Synthesis, Storage, Release and Catabolism

Histamine penetrates the brain poorly from blood, which protects the brain from many effects of blood-borne histamine. Histamine is synthesized from amino acid histidine by the specific histidine decarboxylase (HDC) enzyme in the brain.<sup>2</sup> The activity of HDC is highest in the hypothalamus where the histaminergic cell bodies are located, but HDC is also active in histaminergic nerve terminals.<sup>3</sup> The rate-limiting factor for histamine synthesis is the bioavailability of its precursor, histidine. Histamine is stored in vesicles in cell somata and especially in axon varicosities distinct from those containing GABA in the same cells.<sup>12-14</sup> Vesicular monoamine transporter 2 (VMAT-2) is responsible for the transport of histamine to the intracellular vesicles.<sup>5</sup> Upon arrival of action potentials histamine is released in a Ca<sup>2+</sup>-dependent manner from the storage vesicles. In contrast to other amines, histaminergic synapses are rarely found in vertebrate nervous tissue and most histaminergic endings (varicosities) do not make close contact with postsynaptic sites.

Inactivation of histamine in the brain begins with a methylation reaction by histamine-N-methyltransferase.<sup>15</sup> *Trans*-methylhistamine undergoes oxidative deamination by monoamine oxidase B

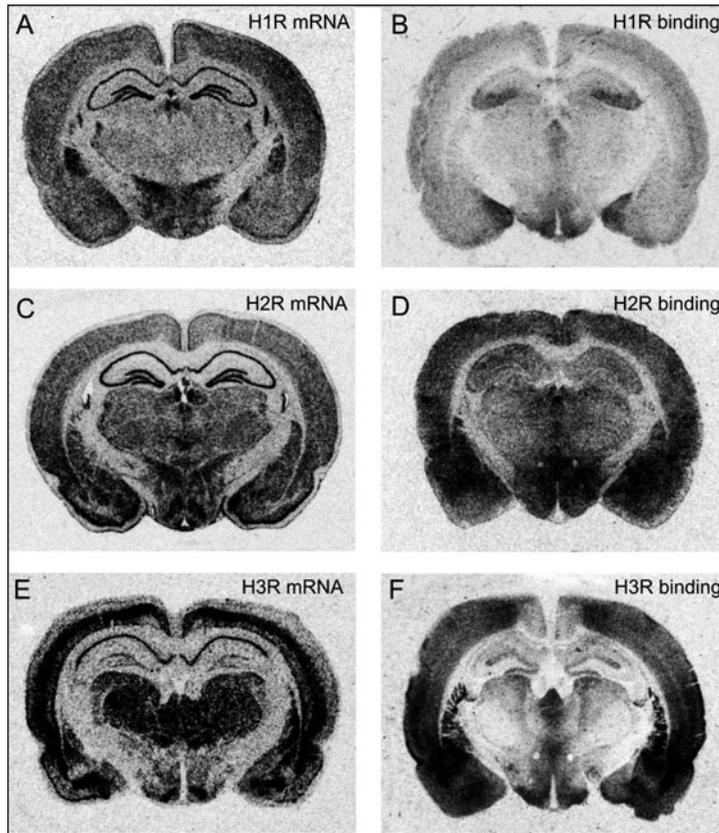


Figure 2. Distribution of histamine receptors in rat brain. Coronal sections at the level of the medial hypothalamus showing mRNA in situ hybridization and specific radioligand binding of H<sub>1</sub>R (A, B), H<sub>2</sub>R (C, D) and H<sub>3</sub>R (E, F) receptors. Modified from: Haas H, Panula P. *Nat Rev Neurosci* 2003; 4:121-130.<sup>5</sup>

(MAO-B) into *t*-methyl-imidazolacetic acid.<sup>16</sup> Diamine oxidase (DAO) is the main histamine metabolizing enzyme in the peripheral tissues, but its activity in the brain is considerably low in basal conditions. However, it can be activated under conditions where methylation is inhibited.<sup>17</sup>

### Histamine Receptors in the Brain

The actions of histamine in the brain are mediated through four G protein-coupled receptors, H<sub>1</sub>R-H<sub>4</sub>R (Fig. 2). H<sub>1</sub>R and H<sub>2</sub>R are postsynaptic and mediate mostly excitatory actions on neurons or potentiate excitatory inputs.<sup>5</sup> H<sub>3</sub>R are located on histaminergic and other neurons on their somata, dendrites and axons.<sup>18</sup> On presynaptic sites H<sub>3</sub> autoreceptors inhibit the synthesis and release of histamine and H<sub>3</sub> heteroreceptors inhibit the release of other neurotransmitters.<sup>19,20</sup> The importance of postsynaptic H<sub>3</sub>R on somata of neurons other than histaminergic neurons is not fully understood, but a recent study demonstrated that in striatum they are able to couple to dopamine D2 receptors and this interaction decreases the affinity of D2 receptors for its agonists.<sup>21</sup> An interesting property of H<sub>3</sub>R is the high constitutive activity, which means spontaneous activity in the absence of histamine.<sup>22</sup> The constitutive activity has a potential regulatory role in the brain and several inverse agonists that are able to block this activity are currently in clinical trials to prove their efficacy in disorders such as Alzheimer's disease, schizophrenia, epilepsy, narcolepsy and obesity.<sup>23,24</sup> Another particular feature of

H<sub>3</sub>R is the existence of multiple isoforms<sup>25,26</sup> derived from a single gene<sup>27</sup> by alternative splicing. The distribution of H<sub>1</sub>R-H<sub>3</sub>R is widespread in the brain as characterized by mRNA in situ hybridization and specific radioligand binding studies (Fig. 2). H<sub>4</sub>R were first thought to be expressed only in the periphery, but recent studies suggest that H<sub>4</sub>R is also expressed both in human and rat brain with highest levels of H<sub>4</sub>R mRNA detected in the spinal cord.<sup>28</sup>

Histamine neurons are pacemakers that display a regular spontaneous firing with low frequency (1-4 Hz). When waking up, the firing of histaminergic neurons is increased.<sup>5</sup> During slow wave sleep the firing is low and no firing can be detected during rapid-eye-movement (REM) sleep. The inhibition of histaminergic neurons during sleep is mediated via GABAergic control from the ventrolateral preoptic area (VLPO).<sup>29</sup>

### ***Physiological Role of Neuronal Histamine***

Brain histamine is implicated in brain homeostasis and control of several neuroendocrine functions. Histamine has an important role in the control of behavioral state, biological rhythms, body weight, energy metabolism, thermoregulation, fluid balance, stress and reproduction.<sup>5</sup> In addition, histamine is implicated in higher brain functions such as sensory and motor functions, mood state, reward, learning and memory.

### **Arousal**

Histamine has a prominent role in the control of arousal. The first cues of histamine as a waking substance came from the unwanted sedative side-effects of the first generation antihistamines that were able to cross the blood brain barrier. EEG recordings have shown that tuberomammillary neurons fire during wakefulness, but not during sleep.<sup>30,31</sup> In agreement, manipulation of the histaminergic system by H<sub>3</sub>R antagonists to activate histaminergic neurons,<sup>32</sup> inhibition of histamine synthesis by alpha-fluoromethylhistidine<sup>33</sup> or histidine decarboxylase gene deletion (HDC knockout mice) leads to disturbances in sleep and waking state.<sup>34</sup>

### **Control of Pituitary Hormone Secretion**

The role of histamine in the regulation of various endocrine functions is due to the effects of histamine on secretion of pituitary hormones.<sup>35</sup> This function is also in agreement with abundant expression of H<sub>1</sub>R-H<sub>3</sub>R in the hypothalamus (Fig. 2). Histamine regulates fluid balance via activation of H<sub>1</sub>R localized on the neurons of supraoptic nucleus, which causes the release of vasopressin<sup>36,37</sup> and in turn induces antidiuresis.<sup>38,39</sup> Histaminergic neurons are also activated during parturition and lactation regulating the release of oxytocin and prolactin.<sup>35,40,41</sup> In addition, certain subgroups of histaminergic neurons are activated in response to stressful stimuli and control the release of adrenocorticotrophic hormone.<sup>42</sup> Histamine also participates in the regulation of the release of growth hormone and thyrotropin-releasing hormone.<sup>35</sup>

### **Appetite and Body Weight**

A large body of evidence links neuronal histamine to the regulation of appetite and body weight. First indications came from the appetite stimulating and weight increasing side-effects of first generation antipsychotics and antidepressants that had strong H<sub>1</sub>R antagonist properties.<sup>43</sup> Later, several studies have demonstrated that histamine acts as an anorexigenic agent via stimulation of H<sub>1</sub>R.<sup>44</sup> Histamine mediates the inhibitory effect of leptin on appetite via H<sub>1</sub>R<sup>45</sup> confirmed by the complete absence of leptin-induced feeding suppression in H<sub>1</sub>R knockout mice.<sup>46,47</sup> The effects of histamine on appetite are linked to various other neuroendocrine peptides such as orexins, neuropeptide Y, peptide YY and bombesin.<sup>48</sup> In addition to control of appetite, neuronal histamine affects metabolism by increasing lipolysis.<sup>49,50</sup> H<sub>3</sub>R are promising targets to treat obesity since blockade of H<sub>3</sub>R seems to be beneficial in decreasing energy intake, body weight and plasma triglycerides.<sup>23</sup> However, based on inconsistent results in H<sub>3</sub>R antagonist studies, further investigations are needed to prove the potential of these drugs in the treatment of obesity and weight gain.

### **Histamine in Brain Diseases**

Even though no pathological states have been selectively connected to deficits in the brain histamine system, alterations in histaminergic neurotransmission have been found in many neurological and psychiatric diseases such as sleep disorders, disorders of mood and cognition (schizophrenia, depression, Alzheimer's disease), movement disorders (Parkinson's disease), epilepsy, eating disorders, pain, neuroinflammation and addiction.<sup>48</sup> In the following parts of this chapter the role of histamine in some selected important brain diseases will be reviewed.

#### **Sleep-Wake Disorders**

Narcolepsy is a rare sleep disorder which is characterized by excessive daytime sleepiness that can be accompanied by manifestation of sudden loss of muscle tone triggered by emotional factors, referred to as cataplexy.<sup>51</sup> Pathophysiological studies have shown that narcolepsy is caused by the early loss of orexinergic neurons in the hypothalamus. Histaminergic neurons remain active during cataplexy whereas norepinephrine neurons stop firing and serotonin neurons lose much of their activity.<sup>52</sup> H<sub>3</sub>R antagonists reduce sleepiness and cataplexy in animal models, probably due to the blockade of autoreceptors on histaminergic neurons resulting in increased release of histamine. Several compounds are being investigated in Phase II clinical trials for the treatment of narcolepsy.<sup>23</sup>

Due to the central role of histamine in the control of arousal and wake state, histamine receptors are potential targets for treatment of other types of sleep and wakefulness disorders as well. Doxepin is a tricyclic antidepressant that displays antagonistic effects on H<sub>1</sub>R/H<sub>2</sub>R in addition to the inhibitory action on norepinephrine and serotonin reuptake. The most common side-effect of doxepin is sedation and it has been shown to improve sleep quality in elderly patients suffering from insomnia.<sup>53</sup> Hypersomnia, on the other hand, could be treated by enhancing histaminergic activity. H<sub>3</sub>R control histaminergic activity and histamine release and thus are promising targets to treat hypersomnia.

#### **Alzheimer's Disease**

Neuropathological studies have demonstrated deficits in the histaminergic system of Alzheimer's disease (AD) patients. Histamine and histidine decarboxylase levels are decreased in some key areas for cognition such as frontal cortex and hippocampus (Table 1).<sup>54,55</sup> Furthermore, numerous neurofibrillary tangles are found in the tuberomammillary nucleus of the AD brain and the number of large neurons in TMN is decreased, which may at least partly cause the histaminergic dysfunction in AD brain.<sup>56,57</sup> The number of H<sub>1</sub>R ligand binding sites is decreased in the AD brain,<sup>58</sup> but interestingly H<sub>3</sub>R levels seem to remain normal.<sup>59</sup> It is possible that decreased histaminergic activity may participate in the cognitive impairments of AD based on the ability of histamine to activate septohippocampal GABAergic neurons through both direct and indirect (cholinergic) mechanisms, which contribute to maintenance of hippocampal theta rhythm and thus cognition and memory.<sup>60</sup> A potentially important target for cognitive effects of H<sub>3</sub>R ligands is also the thalamocortical system since the H<sub>3</sub>R is expressed at particularly high levels in both the rat<sup>61</sup> and human<sup>62</sup> thalamus and cerebral cortex. Interestingly in the mouse, the H<sub>3</sub>R expression is significantly lower, which may render studies with mice less relevant for modeling of human disease processes than those with rats.

Novel H<sub>3</sub>R antagonists increase acetylcholine levels in cortical areas and hippocampus and improve performance in different cognition paradigms in experimental animals.<sup>59,63-65</sup> Ongoing clinical trials will show whether these compounds are effective in patients as well. In addition to memory-improving effects, H<sub>3</sub>R antagonists show efficacy in attention and impulsivity, which makes them attractive candidates for the treatment of attention deficit hyperactivity disorder (ADHD) and cognitive deficits in schizophrenia (see below).

**Table 1. Changes in histamine concentrations in different brain areas of Parkinson's disease and Alzheimer's disease patients**

	Parkinson's Disease	Alzheimer's Disease
Caudate	+28*	±0
Putamen	+59*	-60
Substantia nigra pars compacta	+101*	-27
Substantia nigra pars reticulata	+64	-23
Globus pallidus internum	+134*	-
Globus pallidus externum	+100*	-
Hypothalamus	+47	-58*
Hippocampus	+16	-57*
Frontal cortex	+17	-33
Temporal cortex	-5	-47*
Occipital cortex	-18	-25

Data are expressed as a percentage change in histamine concentration from corresponding control brains. Asterisks refer to statistically significant differences from controls. Modified from Panula et al<sup>54</sup> and Rinne et al.<sup>78</sup>

### Schizophrenia

Studies in humans and models in rodents support a role for histamine in the pathophysiology of schizophrenia. The levels of histamine's major metabolite, N-tele-methylhistamine, are elevated in the cerebrospinal fluid of schizophrenics<sup>66</sup> whereas the H<sub>1</sub>R binding sites are decreased.<sup>67,68</sup> In agreement, repeated administration of methamphetamine which results in behavioral sensitization to dopamine agonists, a cardinal feature of schizophrenia, is accompanied by enhanced histamine release in rat brain.<sup>69</sup> Similar increases in histamine release are found in mice treated with phencyclidine.<sup>70</sup> Histamine H<sub>3</sub>R radioligand binding is significantly increased postmortem in the prefrontal cortex of schizophrenics as compared to normal control, bipolar or depressive subjects.<sup>71</sup> Although this may be a consequence of drug treatment, lack of such differences in the temporal cortex of the same subjects suggests that increased H<sub>3</sub>R radioligand binding may be directly related to the disease process. H<sub>3</sub>R may play important roles in regulation of the thalamocortical system, which is essential for sensory systems and cognitive functions. H<sub>3</sub>R mRNA is expressed at very high level in the dorsal thalamic nuclei of the human brain<sup>72</sup> and in layers IV and V of different subregions of the prefrontal cortex.<sup>62</sup>

Based on the alterations in the histaminergic system in schizophrenic patients, histaminergic drugs might be useful in the treatment of schizophrenia. Indeed, in preclinical studies, H<sub>3</sub>R antagonists display antipsychotic effects by improving the deficits in sensorimotor gating in a prepulse inhibition of startle model and by reducing the hyperactivity induced by methamphetamine.<sup>73</sup> Some new H<sub>3</sub>R antagonists also increase the release of dopamine in rat prefrontal cortex<sup>59,63</sup> which is regarded as beneficial since hypodopaminergic function in prefrontal cortex is associated with negative symptoms and cognitive deficits of schizophrenia.<sup>74</sup> Tiprolisant (BF2.649) is a novel H<sub>3</sub>R antagonist that has an antipsychotic profile in animal models and shows efficacy in patients suffering from antipsychotic-induced weight gain. The effect of tiprolisant on cognitive functions is currently being investigated in clinical trials.<sup>23</sup> Interestingly several open studies have shown that the H<sub>2</sub>R antagonist famotidine reduces negative symptoms in schizophrenics,<sup>75,76</sup> a finding that needs to be confirmed in controlled studies.

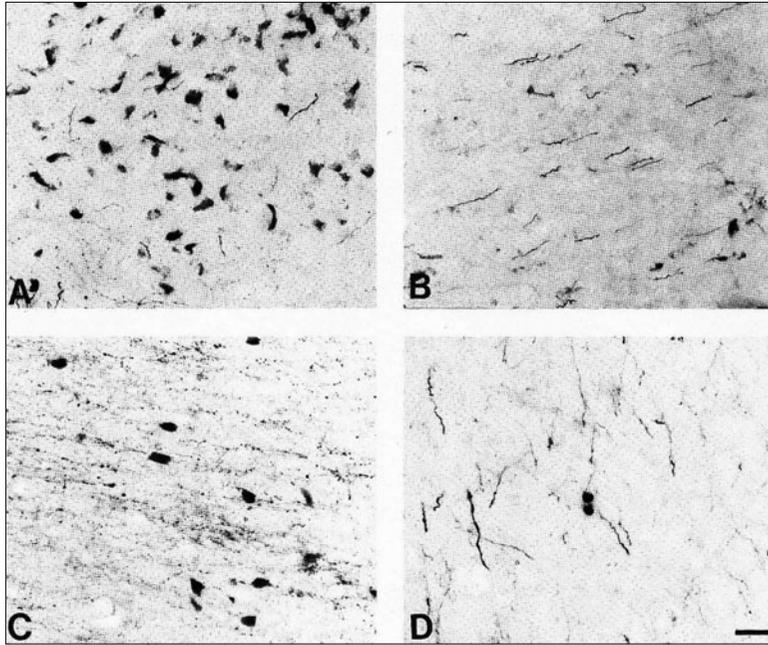


Figure 3. Histaminergic fibers in the substantia nigra of normal and Parkinson's disease brain. A,B) Histamine-immunoreactive fibers in normal brain substantia nigra pars compacta (A) and pars reticulata (B). C,D) Histaminergic fibers in Parkinson's disease brain substantia nigra pars compacta (C) and pars reticulata (D). Reproduced with permission from: Anichtchik OV et al. *Exp Neurol* 2000; 163:20-30,<sup>77</sup> ©2000 Elsevier.

### Parkinson's Disease

Morphological changes and an increase in histaminergic fibers in the substantia nigra have been detected in Parkinson's disease (PD) patients (Fig. 3).<sup>77</sup> Also the histamine levels in the brains of Parkinson patients are increased in areas that regulate motor behavior such as the putamen, substantia nigra and globus pallidus (Table 1).<sup>78</sup> Polymorphism of Thr105Ile histamine-N-methyltransferase that leads to increased turnover of histamine is associated with PD risk.<sup>79</sup> Furthermore, H<sub>3</sub>R ligand binding levels are increased<sup>80</sup> or unchanged<sup>81</sup> in PD patients. The H<sub>3</sub>R binding is also increased in an animal model of PD where dopaminergic neurons are destroyed by a neurotoxic agent 6-OHDA.<sup>82,83</sup> H<sub>3</sub>R regulate the release of GABA<sup>84</sup> and serotonin<sup>85</sup> in the direct and indirect striato-nigral movement pathways and might thus serve as possible drug targets in the treatment of Parkinson's disease and other movement disorders. H<sub>3</sub>R antagonists improved motor coordination in 6-OHDA-lesioned rats.<sup>86</sup> In contrast, the H<sub>3</sub>R agonist immpip, co-administered with L-dopa, decreased dyskinesia in nonhuman primate model of PD, whereas it increased parkinsonian disability when given alone.<sup>87</sup>

Since 1988<sup>88</sup> it has been known that inflammation and microglia may play important roles in PD. Both in animal models of PD and in PD patients, many characteristic features of neuroinflammation have been found.<sup>89</sup> An alteration of blood-brain barrier (BBB) function, characterized by increased permeability to both FITC-labeled albumin and horseradish peroxidase and neovascularization, follows an experimental lesion induced by intracerebral 6-OHDA.<sup>90</sup> Endothelial proliferation has also been found in human PD patients<sup>91</sup> and this seems to be associated with dysfunction of the BBB in PD patients as evidenced by uptake of (<sup>11</sup>C)verapamil.<sup>92</sup> The alteration of BBB function may be a secondary phenomenon, since in experimental animals

it is observed ipsilaterally to the 6-OHDA lesion.<sup>90</sup> Whether the alteration of BBB function is primary or secondary in PD, it is potentially important as dysfunctional BBB may allow entry of blood-borne toxins in the affected areas. Similarly, entrance of histidine might increase leading to larger than normal amounts of substrate available for HDC present in the nerve fibers. Histamine synthesis is largely regulated by substrate availability and unusually high histidine levels following, e.g., portacaval anastomosis are associated with very high brain histamine levels.<sup>93</sup> Alternatively, increased histamine in PD may be associated with neovascularization.

### Epilepsy

Animal models and clinical observations have revealed that the brain histaminergic system has an inhibitory effect on seizures.<sup>5</sup> H<sub>1</sub>R antihistamines show pro-convulsant effects particularly in children<sup>94</sup> and suppression of histaminergic activity promotes seizures in animal models.<sup>95-98</sup> In addition, development of epileptic behavior by kindling of the amygdala is increased in HDC- and H<sub>1</sub>R-knockout mice.<sup>99</sup> Histamine levels in cerebrospinal fluid of children with febrile convulsions are significantly lower than those in the febrile children without convulsions.<sup>100</sup> Low histamine levels have also been detected in Krushinski-Molodkina rats that are prone to epilepsy.<sup>101</sup> In addition to its anti-convulsive effects, histamine has neuroprotective properties<sup>102,103</sup> and inhibits excitotoxic effects of glutamate.<sup>103</sup> H<sub>3</sub>R antagonists have shown beneficial effects in different seizure models, such as electrically induced convulsions,<sup>104,105</sup> kindling<sup>106</sup> and pentylenetetrazole-induced seizures.<sup>107</sup>

### Brain Injury

Involvement of histamine in the pathophysiology of brain injuries has been demonstrated in hypoxia,<sup>108</sup> trauma,<sup>109,110</sup> ischemia and stroke<sup>111</sup> or neoplasms.<sup>112</sup> For example, in rat brain, trauma leads to increased histamine levels both in the plasma and in the traumatized hemisphere.<sup>110</sup> In another study, fluid-percussion-evoked brain trauma caused changes in H<sub>3</sub>R regulation demonstrated by receptor binding and mRNA in situ hybridization experiments.<sup>109</sup> In ischemic brain damage, histamine is released gradually over a long time frame.<sup>113</sup> Since preischemic administration of alpha-fluoromethylhistidine completely abolishes the ischemia-induced increase in the brain histamine, the source of histamine in cerebral ischemia is regarded to be neuronal.<sup>113,114</sup> An increased level of brain histamine may contribute to the amelioration of ischemic neuronal damage.<sup>115</sup> Thus, the emergence of neuronal injuries after ischemic events could be decreased by increasing brain histamine levels by histidine loading or H<sub>3</sub>R blockade.

### Pain

The descending histaminergic neurons originating from TMN project to areas related to pain perception<sup>116</sup> such as the dorsal raphe nucleus, periaqueductal gray region and dorsal horn of the spinal cord.<sup>4,117,118</sup> Several studies have demonstrated that histamine applied directly into CNS induces antinociception.<sup>119-123</sup> In line with this, brain histamine level reduction by alpha-fluoromethylhistidine or H<sub>3</sub>R agonists enhances nociception.<sup>123,124</sup> Studies using histaminergic ligands<sup>121</sup> or mice lacking either H<sub>1</sub>R or H<sub>2</sub>R<sup>125</sup> suggest that H<sub>1</sub>R and H<sub>2</sub>R are responsible for the mediation of histamine's effects on pain perception.

Spinal H<sub>3</sub>R have been also linked to antinociception based on studies with H<sub>3</sub>R knockout mice and H<sub>3</sub>R antagonists.<sup>126,127</sup> Interestingly, activation of H<sub>3</sub>R seems to inhibit only certain types of pain such as pain induced by mechanical low-intensity stimulation.<sup>128</sup> The antinociceptive properties of novel H<sub>4</sub>R antagonists<sup>129</sup> and recently described expression of H<sub>4</sub>R in spinal cord<sup>128</sup> suggest a role for neuronal H<sub>4</sub>R in pain perception as well.

### Conclusion

The brain histaminergic system participates in the regulation of various brain functions, including sleep-wake cycle, energy and endocrine homeostasis and cognition. The effects of histamine in the brain are mediated via four histamine receptors (H<sub>1</sub>R-H<sub>4</sub>R), of which H<sub>1</sub>R-H<sub>3</sub>R are highly expressed. Changes in the neuronal histaminergic system are found in various brain

disorders such as Alzheimer's disease, Parkinson's disease and schizophrenia, making histamine receptors promising targets for future drug therapies.

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### References

1. Abel JJ, Kubota S. On the presence of histamine (-iminazolyethylamine) in the hypophysis cerebri and other tissues of the body and its occurrence among the hydrolytic decomposition products of proteins. *J Pharmacol Exp Ther* 1919; 13:243-300.
2. Garbarg M, Barbin G, Bischoff S et al. Evidence for a specific decarboxylase involved in histamine synthesis in an ascending pathway in rat brain. *Agents Actions* 1974; 4:181.
3. Watanabe T et al. Evidence for the presence of a histaminergic neuron system in the rat brain: an immunohistochemical analysis. *Neurosci Lett* 1983; 39:249-254.
4. Panula P, Yang HY, Costa E. Histamine-containing neurons in the rat hypothalamus. *Proc Natl Acad Sci USA* 1984; 81:2572-2576.
5. Haas H, Panula P. The role of histamine and the tuberomammillary nucleus in the nervous system. *Nat Rev Neurosci* 2003; 4:121-130.
6. Ericson H, Watanabe T, Kohler C. Morphological analysis of the tuberomammillary nucleus in the rat brain: delineation of subgroups with antibody against L-histidine decarboxylase as a marker. *J Comp Neurol* 1987; 263:1-24.
7. Panula P, Airaksinen MS, Pirvola U et al. A histamine-containing neuronal system in human brain. *Neuroscience* 1990; 34:127-132.
8. Martres MP, Baudry M, Schwartz JC. Histamine synthesis in the developing rat brain: evidence for a multiple compartmentation. *Brain Res* 1975; 83:261-275.
9. Katoh Y et al. Histamine production by cultured microglial cells of the mouse. *Neurosci Lett* 2001; 305:181-184.
10. Karlstedt K, Nissinen M, Michelsen KA et al. Multiple sites of L-histidine decarboxylase expression in mouse suggest novel developmental functions for histamine. *Dev Dyn* 2001; 221:81-91.
11. Molina-Hernandez A, Velasco I. Histamine induces neural stem cell proliferation and neuronal differentiation by activation of distinct histamine receptors. *J Neurochem* 2008; 106:706-717.
12. Kukko-Lukjanov TK, Panula P. Subcellular distribution of histamine, GABA and galanin in tuberomammillary neurons in vitro. *J Chem Neuroanat* 2003; 25:279-292.
13. Erickson JD, Schafer MK, Bonner TI et al. Distinct pharmacological properties and distribution in neurons and endocrine cells of two isoforms of the human vesicular monoamine transporter. *Proc Natl Acad Sci USA* 1996; 93:5166-5171.
14. Merickel A, Edwards RH. Transport of histamine by vesicular monoamine transporter-2. *Neuropharmacology* 1995; 34:1543-1547.
15. Reilly MA, Schayer RW. In vivo studies on histamine catabolism and its inhibition. *Br J Pharmacol* 1970; 38:478-489.
16. Hough LB, Domino EF. Tele-methylhistamine oxidation by type B monoamine oxidase. *J Pharmacol Exp Ther* 1979; 208:422-428.
17. Prell GD, Morrishow AM, Duoyon E et al. Inhibitors of histamine methylation in brain promote formation of imidazoleacetic acid, which interacts with GABA receptors. *J Neurochem* 1997; 68:142-151.
18. Arrang JM, Garbarg M, Schwartz JC. Auto-inhibition of brain histamine release mediated by a novel class (H3) of histamine receptor. *Nature* 1983; 302:832-837.
19. Schlicker E, Malinowska B, Kathmann M et al. Modulation of neurotransmitter release via histamine H3 heteroreceptors. *Fundam Clin Pharmacol* 1994; 8:128-137.
20. Brown RE, Stevens DR, Haas HL. The physiology of brain histamine. *Prog Neurobiol* 2001; 63:637-672.
21. Ferrada C et al. Interactions between histamine H3 and dopamine D2 receptors and the implications for striatal function. *Neuropharmacology* 2008; 55:190-197.
22. Morisset S et al. High constitutive activity of native H3 receptors regulates histamine neurons in brain. *Nature* 2000; 408:860-864.
23. Sander K, Kottke T, Stark H. Histamine H3 receptor antagonists go to clinics. *Biol Pharm Bull* 2008; 31:2163-2181.
24. Arrang JM, Morisset S, Gbahou F. Constitutive activity of the histamine H3 receptor. *Trends Pharmacol Sci* 2007; 28:350-357.
25. Drutel G et al. Identification of rat H3 receptor isoforms with different brain expression and signaling properties. *Mol Pharmacol* 2001; 59:1-8.

26. Rouleau A et al. Cloning and expression of the mouse histamine H3 receptor: evidence for multiple isoforms. *J Neurochem* 2004; 90:1331-1338.
27. Lovenberg TW et al. Cloning and functional expression of the human histamine H3 receptor. *Mol Pharmacol* 1999; 55:1101-1107.
28. Strakhova MI et al. Localization of histamine H(4) receptors in the central nervous system of human and rat. *Brain Res* 2008.
29. Sherin JE, Shiromani PJ, McCarley RW et al. Activation of ventrolateral preoptic neurons during sleep. *Science* 1996; 271:216-219.
30. Lin JS, Hou Y, Sakai K et al. Histaminergic descending inputs to the mesopontine tegmentum and their role in the control of cortical activation and wakefulness in the cat. *J Neurosci* 1996; 16:1523-1537.
31. Lin JS. Brain structures and mechanisms involved in the control of cortical activation and wakefulness, with emphasis on the posterior hypothalamus and histaminergic neurons. *Sleep Med Rev* 2000; 4:471-503.
32. Lin JS et al. Involvement of histaminergic neurons in arousal mechanisms demonstrated with H3-receptor ligands in the cat. *Brain Res* 1990; 523:325-330.
33. Kiyono S et al. Effects of alpha-fluoromethylhistidine on sleep-waking parameters in rats. *Physiol Behav* 1985; 34:615-617.
34. Parmentier R et al. Anatomical, physiological and pharmacological characteristics of histidine decarboxylase knock-out mice: evidence for the role of brain histamine in behavioral and sleep-wake control. *J Neurosci* 2002; 22:7695-7711.
35. Knigge U, Warberg J. The role of histamine in the neuroendocrine regulation of pituitary hormone secretion. *Acta Endocrinol (Copenh)* 1991; 124:609-619.
36. Armstrong WE, Sladek CD. Evidence for excitatory actions of histamine on supraoptic neurons in vitro: mediation by an H1-type receptor. *Neuroscience* 1985; 16:307-322.
37. Haas HL, Wolf P, Nussbaumer JC. Histamine: action on supraoptic and other hypothalamic neurones of the cat. *Brain Res* 1975; 88:166-170.
38. Bhargava KP, Kulshrestha VK, Santhakumari G et al. Mechanism of histamine-induced antidiuretic response. *Br J Pharmacol* 1973; 47:700-706.
39. Tuomisto L, Eriksson L, Fyhrquist F. Vasopressin release by histamine in the conscious goat. *Eur J Pharmacol* 1980; 63:15-24.
40. Libertun C, McCann SM. The possible role of histamine in the control of prolactin and gonadotropin release. *Neuroendocrinology* 1976; 20:110-120.
41. Hashimoto H, Noto T, Nakajima T. A study on the release mechanism of vasopressin and oxytocin. *Neuropeptides* 1988; 12:199-206.
42. Miklos IH, Kovacs KJ. Functional heterogeneity of the responses of histaminergic neuron subpopulations to various stress challenges. *Eur J Neurosci* 2003; 18:3069-3079.
43. Kalucy RS. Drug-induced weight gain. *Drugs* 1980; 19:268-278.
44. Jorgensen EA, Knigge U, Warberg J et al. Histamine and the regulation of body weight. *Neuroendocrinology* 2007; 86:210-214.
45. Morimoto T et al. Involvement of the histaminergic system in leptin-induced suppression of food intake. *Physiol Behav* 1999; 67:679-683.
46. Masaki T, Yoshimatsu H, Chiba S et al. Targeted disruption of histamine H1-receptor attenuates regulatory effects of leptin on feeding, adiposity and UCP family in mice. *Diabetes* 2001; 50:385-391.
47. Masaki T et al. Involvement of hypothalamic histamine H1 receptor in the regulation of feeding rhythm and obesity. *Diabetes* 2004; 53:2250-2260.
48. Haas HL, Sergeeva OA, Selbach O. Histamine in the nervous system. *Physiol Rev* 2008; 88:1183-1241.
49. Bugajski J, Janusz Z. Lipolytic responses induced by intracerebroventricular administration of histamine in the rat. *Agents Actions* 1981; 11:147-150.
50. Yoshimatsu H et al. Histidine induces lipolysis through sympathetic nerve in white adipose tissue. *Eur J Clin Invest* 2002; 32:236-241.
51. Dauvilliers Y, Arnulf I, Mignot E. Narcolepsy with cataplexy. *Lancet* 2007; 369:499-511.
52. John J, Wu MF, Boehmer LN et al. Cataplexy-active neurons in the hypothalamus: implications for the role of histamine in sleep and waking behavior. *Neuron* 2004; 42:619-634.
53. Scharf M et al. Efficacy and Safety of Doxepin 1 mg, 3 mg and 6 mg in Elderly Patients With Primary Insomnia: a Randomized, Double-Blind, Placebo-Controlled Crossover Study. *J Clin Psychiatry* 2008.
54. Panula P et al. Neuronal histamine deficit in Alzheimer's disease. *Neuroscience* 1998; 82:993-997.
55. Mazurkiewicz-Kwilecki IM, Nsonwah S. Changes in the regional brain histamine and histidine levels in postmortem brains of Alzheimer patients. *Can J Physiol Pharmacol* 1989; 67:75-78.
56. Nakamura S et al. Loss of large neurons and occurrence of neurofibrillary tangles in the tuberomammillary nucleus of patients with Alzheimer's disease. *Neurosci Lett* 1993; 151:196-199.
57. Airaksinen MS, Reinikainen K, Riekkinen P et al. Neurofibrillary tangles and histamine-containing neurons in Alzheimer hypothalamus. *Agents Actions* 1991; 33:104-107.

58. Higuchi M et al. Histamine H(1) receptors in patients with Alzheimer's disease assessed by positron emission tomography. *Neuroscience* 2000; 99:721-729.
59. Medhurst AD et al. GSK189254, a novel H3 receptor antagonist that binds to histamine H3 receptors in Alzheimer's disease brain and improves cognitive performance in preclinical models. *J Pharmacol Exp Ther* 2007; 321:1032-1045.
60. Xu C et al. Histamine innervation and activation of septohippocampal GABAergic neurons: involvement of local ACh release. *J Physiol* 2004; 561:657-670.
61. Jin C, Lintunen M, Panula P. Histamine H(1) and H(3) receptors in the rat thalamus and their modulation after systemic kainic acid administration. *Exp Neurol* 2005; 194:43-56.
62. Jin CY, Panula P. The laminar histamine receptor system in human prefrontal cortex suggests multiple levels of histaminergic regulation. *Neuroscience* 2005; 132:137-149.
63. Ligneau X et al. BF2.649 (1-{3-(3-(4-Chlorophenyl)propoxy)propyl}piperidine, hydrochloride), a nonimidazole inverse agonist/antagonist at the human histamine H3 receptor: preclinical pharmacology. *J Pharmacol Exp Ther* 2007; 320:365-375.
64. Fox GB et al. Pharmacological properties of ABT-239 (4-(2-{2-((2R)-2-Methylpyrrolidinyl)ethyl}-benzofuran-5-yl)benzonitrile): II. Neurophysiological characterization and broad preclinical efficacy in cognition and schizophrenia of a potent and selective histamine H3 receptor antagonist. *J Pharmacol Exp Ther* 2005; 313:176-190.
65. Cowart M et al. 4-(2-(2-(R)-methylpyrrolidin-1-yl)ethyl)benzofuran-5-yl)benzonitrile and related 2-aminoethylbenzofuran H3 receptor antagonists potently enhance cognition and attention. *J Med Chem* 2005; 48:38-55.
66. Prell GD et al. Histamine metabolites in cerebrospinal fluid of patients with chronic schizophrenia: their relationships to levels of other aminergic transmitters and ratings of symptoms. *Schizophr Res* 1995; 14:93-104.
67. Nakai T et al. Decreased histamine H1 receptors in the frontal cortex of brains from patients with chronic schizophrenia. *Biol Psychiatry* 1991; 30:349-356.
68. Iwabuchi K et al. Histamine H1 receptors in schizophrenic patients measured by positron emission tomography. *Eur Neuropsychopharmacol* 2005; 15:185-191.
69. Morisset S et al. Acute and chronic effects of methamphetamine on tele-methylhistamine levels in mouse brain: selective involvement of the D(2) and not D(3) receptor. *J Pharmacol Exp Ther* 2002; 300:621-628.
70. Itoh Y, Oishi R, Nishibori M et al. Phencyclidine and the dynamics of mouse brain histamine. *J Pharmacol Exp Ther* 1985; 235:788-792.
71. Jin C, Anichtchik O, Panula P. Altered histamine H3 receptor radioligand binding in postmortem brain samples from subjects with psychiatric diseases. *Br J Pharmacol*, In press 2009.
72. Jin CY, Kalimo H, Panula P. The histaminergic system in human thalamus: correlation of innervation to receptor expression. *Eur J Neurosci* 2002; 15:1125-1138.
73. Browman KE et al. Enhancement of prepulse inhibition of startle in mice by the H3 receptor antagonists thioperamide and ciproxifan. *Behav Brain Res* 2004; 153:69-76.
74. Esbenshade TA et al. The histamine H3 receptor: an attractive target for the treatment of cognitive disorders. *Br J Pharmacol* 2008; 154:1166-1181.
75. Kaminsky R, Moriarty TM, Bodine J et al. Effect of famotidine on deficit symptoms of schizophrenia. *Lancet* 1990; 335:1351-1352.
76. Martinez MC. Famotidine in the management of schizophrenia. *Ann Pharmacother* 1999; 33:742-747.
77. Anichtchik OV, Rinne JO, Kalimo H et al. An altered histaminergic innervation of the substantia nigra in Parkinson's disease. *Exp Neurol* 2000; 163:20-30.
78. Rinne JO et al. Increased brain histamine levels in Parkinson's disease but not in multiple system atrophy. *J Neurochem* 2002; 81:954-960.
79. Agundez JA et al. Nonsynonymous polymorphisms of histamine-metabolising enzymes in patients with Parkinson's disease. *Neuromolecular Med* 2008; 10:10-16.
80. Anichtchik OV, Peitsaro N, Rinne JO et al. Distribution and modulation of histamine H(3) receptors in basal ganglia and frontal cortex of healthy controls and patients with Parkinson's disease. *Neurobiol Dis* 2001; 8:707-716.
81. Goodchild RE et al. Distribution of histamine H3-receptor binding in the normal human basal ganglia: comparison with Huntington's and Parkinson's disease cases. *Eur J Neurosci* 1999; 11:449-456.
82. Ryu JH, Yanai K, Watanabe T. Marked increase in histamine H3 receptors in the striatum and substantia nigra after 6-hydroxydopamine-induced denervation of dopaminergic neurons: an autoradiographic study. *Neurosci Lett* 1994; 178:19-22.
83. Anichtchik OV et al. Modulation of histamine H3 receptors in the brain of 6-hydroxydopamine-lesioned rats. *Eur J Neurosci* 2000; 12:3823-3832.

84. Garcia M, Floran B, Arias-Montano JA et al. Histamine H3 receptor activation selectively inhibits dopamine D1 receptor-dependent (3H)GABA release from depolarization-stimulated slices of rat substantia nigra pars reticulata. *Neuroscience* 1997; 80:241-249.
85. Threlfell S et al. Histamine H3 receptors inhibit serotonin release in substantia nigra pars reticulata. *J Neurosci* 2004; 24:8704-8710.
86. Nowak P et al. Histamine H(3) receptor ligands modulate L-dopa-evoked behavioral responses and L-dopa derived extracellular dopamine in dopamine-denervated rat striatum. *Neurotox Res* 2008; 13:231-240.
87. Gomez-Ramirez J, Johnston TH, Visanji NP et al. Histamine H3 receptor agonists reduce L-dopa-induced chorea, but not dystonia, in the MPTP-lesioned nonhuman primate model of Parkinson's disease. *Mov Disord* 2006; 21:839-846.
88. McGeer PL, Itagaki S, Boyes BE et al. Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains. *Neurology* 1988; 38:1285-1291.
89. Whitton PS. Inflammation as a causative factor in the aetiology of Parkinson's disease. *Br J Pharmacol* 2007; 150:963-976.
90. Carvey PM et al. 6-Hydroxydopamine-induced alterations in blood-brain barrier permeability. *Eur J Neurosci* 2005; 22:1158-1168.
91. Faucheux BA, Bonnet AM, Agid Y et al. Blood vessels change in the mesencephalon of patients with Parkinson's disease. *Lancet* 1999; 353:981-982.
92. Kortekaas R et al. Blood-brain barrier dysfunction in parkinsonian midbrain in vivo. *Ann Neurol* 2005; 57:176-179.
93. Fogel WA et al. Neuronal storage of histamine in the brain and tele-methylimidazoleacetic acid excretion in portocaval shunted rats. *J Neurochem* 2002; 80:375-382.
94. Haruyama W et al. The relationship between drug treatment and the clinical characteristics of febrile seizures. *World J Pediatr* 2008; 4:202-205.
95. Yokoyama H et al. Histamine levels and clonic convulsions of electrically-induced seizure in mice: the effects of alpha-fluoromethylhistidine and metoprine. *Naunyn Schmiedebergs Arch Pharmacol* 1992; 346:40-45.
96. Chen Z, Li WD, Zhu LJ et al. Effects of histidine, a precursor of histamine, on pentylentetrazole-induced seizures in rats. *Acta Pharmacol Sin* 2002; 23:361-366.
97. Fujii Y, Tanaka T, Harada C et al. Epileptogenic activity induced by histamine H(1) antagonists in amygdala-kindled rats. *Brain Res* 2003; 991:258-261.
98. Yokoyama H, Sato M, Iinuma K et al. Centrally acting histamine H1 antagonists promote the development of amygdala kindling in rats. *Neurosci Lett* 1996; 217:194-196.
99. Hirai T et al. Development of amygdaloid kindling in histidine decarboxylase-deficient and histamine H1 receptor-deficient mice. *Epilepsia* 2004; 45:309-313.
100. Kiviranta T, Tuomisto L, Airaksinen EM. Histamine in cerebrospinal fluid of children with febrile convulsions. *Epilepsia* 1995; 36:276-280.
101. Onodera K, Tuomisto L, Tacke U et al. Strain differences in regional brain histamine levels between genetically epilepsy-prone and resistant rats. *Methods Find Exp Clin Pharmacol* 1992; 14:13-16.
102. Kukko-Lukjanov TK et al. Histaminergic neurons protect the developing hippocampus from kainic acid-induced neuronal damage in an organotypic coculture system. *J Neurosci* 2006; 26:1088-1097.
103. Dai H et al. Histamine protects against NMDA-induced necrosis in cultured cortical neurons through H receptor/cyclic AMP/protein kinase A and H receptor/GABA release pathways. *J Neurochem* 2006; 96:1390-1400.
104. Yokoyama H, Onodera K, Iinuma K et al. Effect of thioperamide, a histamine H3 receptor antagonist, on electrically induced convulsions in mice. *Eur J Pharmacol* 1993; 234:129-133.
105. Yokoyama H et al. Clobenpropit (VUF-9153), a new histamine H3 receptor antagonist, inhibits electrically induced convulsions in mice. *Eur J Pharmacol* 1994; 260:23-28.
106. Harada C et al. Intracerebroventricular administration of histamine H3 receptor antagonists decreases seizures in rat models of epilepsy. *Methods Find Exp Clin Pharmacol* 2004; 26:263-270.
107. Vohora D, Pal SN, Pillai KK. Histamine and selective H3-receptor ligands: a possible role in the mechanism and management of epilepsy. *Pharmacol Biochem Behav* 2001; 68:735-741.
108. Dux E et al. The blood-brain barrier in hypoxia: ultrastructural aspects and adenylate cyclase activity of brain capillaries. *Neuroscience* 1984; 12:951-958.
109. Lozada A, Maegele M, Stark H et al. Traumatic brain injury results in mast cell increase and changes in regulation of central histamine receptors. *Neuropathol Appl Neurobiol* 2005; 31:150-162.
110. Mohanty S et al. Role of histamine in traumatic brain edema. An experimental study in the rat. *J Neurol Sci* 1989; 90:87-97.
111. Waskiewicz J, Molchanova L, Walajcys-Rode E et al. Hypoxia and ischemia modifies histamine metabolism and transport in brain synaptosomes. *Resuscitation* 1988; 16:287-293.

112. Lefranc F, Yeaton P, Brotchi J et al. Cimetidine, an unexpected anti-tumor agent and its potential for the treatment of glioblastoma (review). *Int J Oncol* 2006; 28:1021-1030.
113. Adachi N, Itoh Y, Oishi R et al. Direct evidence for increased continuous histamine release in the striatum of conscious freely moving rats produced by middle cerebral artery occlusion. *J Cereb Blood Flow Metab* 1992; 12:477-483.
114. Adachi N, Oishi R, Saeki K. Changes in the metabolism of histamine and monoamines after occlusion of the middle cerebral artery in rats. *J Neurochem* 1991; 57:61-66.
115. Adachi N. Cerebral ischemia and brain histamine. *Brain Res Brain Res Rev* 2005; 50:275-286.
116. Basbaum AI, Fields HL. Endogenous pain control systems: brainstem spinal pathways and endorphin circuitry. *Annu Rev Neurosci* 1984; 7:309-338.
117. Panula P et al. Histamine-immunoreactive nerve fibers in the mammalian spinal cord. *Brain Res* 1989; 484:234-239.
118. Panula P, Pirvola U, Auvinen S et al. Histamine-immunoreactive nerve fibers in the rat brain. *Neuroscience* 1989; 28:585-610.
119. Parolaro D et al. Histamine as a central modulator of rat intestinal transit. *J Pharmacol Exp Ther* 1989; 249:324-328.
120. Bhattacharya SK, Parmar SS. Antinociceptive effect of intracerebroventricularly administered histamine in rats. *Res Commun Chem Pathol Pharmacol* 1985; 49:125-136.
121. Thoburn KK, Hough LB, Nalwalk JW et al. Histamine-induced modulation of nociceptive responses. *Pain* 1994; 58:29-37.
122. Glick SD, Crane LA. Opiate-like and abstinence-like effects of intracerebral histamine administration in rats. *Nature* 1978; 273:547-549.
123. Malmberg-Aiello P, Lamberti C, Ghelardini C et al. Role of histamine in rodent antinociception. *Br J Pharmacol* 1994; 111:1269-1279.
124. Malmberg-Aiello P, Lamberti C, Ipponi A et al. Evidence for hypernociception induction following histamine H1 receptor activation in rodents. *Life Sci* 1998; 63:463-476.
125. Mobarakeh JI et al. Enhanced antinociception by intracerebroventricularly administered orexin A in histamine H1 or H2 receptor gene knockout mice. *Pain* 2005; 118:254-262.
126. Cannon KE, Leurs R, Hough LB. Activation of peripheral and spinal histamine H3 receptors inhibits formalin-induced inflammation and nociception, respectively. *Pharmacol Biochem Behav* 2007; 88:122-129.
127. Cannon KE et al. Activation of spinal histamine H3 receptors inhibits mechanical nociception. *Eur J Pharmacol* 2003; 470:139-147.
128. Cannon KE, Hough LB. Inhibition of chemical and low-intensity mechanical nociception by activation of histamine H3 receptors. *J Pain* 2005; 6:193-200.
129. Coruzzi G, Adami M, Guaita E et al. Anti-inflammatory and antinociceptive effects of the selective histamine H4-receptor antagonists JNJ777120 and VUF6002 in a rat model of carrageenan-induced acute inflammation. *Eur J Pharmacol* 2007; 563:240-244.